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Mutated gene coding for a LAT protein  
and the biological applications thereof.

The present invention relates to a mutated gene coding for two mutant LAT proteins leading to an exaggerated T<sub>H</sub>2 cell differentiation. The invention relates to biological structures containing said mutant, particularly, non-human 5 LAT gene mutated animals, cell cultures, plasmids, chromosomal DNAs, embryos comprising said mutated gene, and applications thereof. The invention further relates to screening methods for drug useful for treatment against asthma, allergy and any pathological immune responses 10 involving T<sub>H</sub>2 cells. The invention also relates to method for producing IgE antibodies.

#### **Background Art**

A key event in the pathogenesis is the production of 15 antibodies of the IgE class. Hypergammaglobulinemia E results from loss of immunoregulation. More specifically, T lymphocyte abnormalities have been reported in a number of pathologic hyper IgE conditions and are the object of much research aiming at developing pharmaceutical compounds that 20 will prevent atopic allergy and asthma.

TCR recognize peptide fragments bound to major histocompatibility complex (MHC) molecules and relay this information to the interior of the T cell via adapter 25 proteins. One of these, the adapter LAT (Linker for Activation of T cells), coordinates the assembly of signaling complexes through multiple tyrosine residues within its intracytoplasmic segment. Upon TCR-induced phosphorylation, each of these tyrosine residues manifests 30 some specialization in the signaling proteins it recruits. Studies on cell lines showed that mutation of tyrosine 136 (Y136) selectively eliminates binding of phospholipase C $\gamma$ 1 (PLC- $\gamma$ 1) whereas the simultaneous mutation of Y175, Y195 and

Y235 results in loss of binding of downstream adapters Gads and Grb-2 (Lin and Weiss, 2001; Samelson et al, 1999 ; Zhang et al, 2000). Studies of LAT "knock in" mutant mice presenting the mutation of the four distal tyrosine residues 5 of LAT in phenylalanine, called 4YF mice, showed that the murine T cell development was completely blocked (Sommers et al, 2001). Hence, their thymocyte development was arrested at the immature CD4<sup>+</sup> CD8<sup>-</sup> stage and no mature T cells were present.

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The present invention now provides genetic evidence that LAT exerts an unanticipated and surprising inhibitory function on the differentiation of CD4 helper T (T<sub>H</sub>) cells into T<sub>H2</sub> cells.

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Mice homozygous for the mutation of a single LAT tyrosine (LAT Y136F) results in mice that show a precocious and spontaneous accumulation of polyclonal T<sub>H2</sub> cells, which chronically produce large amounts of interleukins 4, 5, 10 20 and 13. This exaggerated T<sub>H2</sub> differentiation leads in turn to tissue eosinophilia and to the maturation of massive numbers of plasma cells secreting IgE and IgG1 antibodies (see Figure 1). Thus, in addition to known positive signaling, LAT also appears essential for establishing inhibitory 25 signals that control T cell homeostasis.

Mice for the composite mutation of the three distal LAT tyrosines (LAT Y175F+Y195F+Y235F) prevents the development 30 of T cells expressing alpha/beta T cell receptor. However, it allows the development of T cells expressing gamma/delta T cell receptors, and their accumulation in the periphery (see figure 9). These polyclonal gamma/delta T cells chronically produce large amounts of interleukins 4, 5, 10 and 13 (i.e. they present blatant TH2 phenotype). This 35 exaggerated T<sub>H2</sub>-type differentiation of gamma/delta T cells leads in turn to the maturation of massive numbers of plasma cells secreting IgE and IgG1 antibodies (see Figures 10 and

11).

#### Description of drawings

Figure 1 is a diagram disclosing the immune system  
5 development of mutant mice.

Figure 2 illustrates the LAT Y136F knock-in strategy:

(1) : the partial restriction map of the wild-type LAT  
gene.

(2) : the targeting vector used for the introduction of  
10 the LAT Y136F mutation.

(3) : the structure of the targeted allele following  
homologous recombination.

(4) : the final structure of the targeted allele after  
removal of the  $neo^r$  gene via Cre-mediated recombination.

15 Figure 3 illustrates the aberrant growth of lymphoid organs  
in the mice : thymus (A), spleen (B) and lymph nodes (C).

Figure 4 relates to constitutive type-2 cytokine production  
in CD4 T cells freshly isolated from  $LAT^{Y136F}$  peripheral  
lymphoid organs.

20 Figure 5 relates to a phenotypic analysis of T cells from  
wild-type and  $LAT^{Y136F}$  mice.

Figure 6 illustrates eosinophilia in 6 weeks old LATY136F  
lymphoid organs.

25 A: Dot plot panel showing the gate selected for the  
analysis described in panel B and for the sorted cells  
picture in panel C.

B: Single color histograms of gated cells labelled with  
antibodies characterizing eosinophils.

C: Hematoxylin and eosin staining of sorted cells.

30 Figure 7 illustrates the hyperactivity of B lymphocytes:  
massive serum levels of IgE and IgG1 antibodies in  
unimmunized  $LAT^{Y136F}$  mice.

Figure 8 illustrates the LAT Y175F+Y195F+Y235F knock in  
strategy:

35 (1) : the partial restriction map of the wild-type LAT  
gene.

(2) : the targeting vector used for the introduction of

the LAT Y175F, Y195F and Y235F mutation.

(3) : the structure of the targeted allele following homologous recombination.

(4) : the final structure of the targeted allele after removal of the neo<sup>r</sup> gene via Cre-mediated recombination.

Figure 9 relates to a phenotypic analysis of the gamma/delta T cells developed in large numbers in the LAT Y175F+Y195F+Y235F mutant in the mere absence of alpha/beta T cells.

Figure 10 illustrates the T<sub>H</sub>2-type cytokines that are spontaneously produced by the gamma/delta T cells present in LAT Y175F+Y195F+Y235F mutant mice (lane 3) and compare them the T<sub>H</sub>2-type cytokines that are spontaneously produced by the alpha/beta T cells developed in the LAT Y136F mutant (lane 1).

Figure 11 illustrates the hyperactivity of B lymphocytes and the massive amounts of IgE and IgG1 that are spontaneously found in the serum of unimmunized LAT Y175F+Y195F+Y235F mice.

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#### **Description**

In this application, LAT Y136F, LAT Y175F, LAT Y195F, and LAT Y235F refer to the designated mutation itself, while LAT<sup>Y136F</sup>, LAT<sup>Y175F</sup>, LAT<sup>Y195F</sup> and LAT<sup>Y235F</sup> refer to mutants, mice 25 or products derived from these mutations.

Mutation of one or three tyrosine(s) among the four distal tyrosine of LAT protein (i.e. LAT Y136F, or LAT Y175F+Y195F+Y235F) is able to induce the development of 30 pathologies associated with exacerbated T<sub>H</sub>2 immunity. Characteristics of the phenotype associated with this mutation are described in the following examples. Therefore, the present invention provides models of allergy and/or asthma or other diseases associated with T<sub>H</sub>2 cell 35 deregulation or activity, more particularly T<sub>H</sub>2 cell accumulation. Among the advantages of said models, it is found the rapidity of the model preparation (about 3-4 weeks

for a mice model instead of several months) and the exacerbated phenotype (for instance, exacerbated IgE production and tissue eosinophilia).

5 This phenotype due to the mutation of one or three tyrosine(s) among the four distal tyrosine residues of LAT protein (namely, LAT Y136F, or LAT Y175F+Y195F+Y235F) in mice was unpredictable, considering the phenotype of mice in which the four mutations are combined (LAT 4YF mice).  
10 Indeed, LAT 4YF mice are totally devoid of thymocytes and T cells, because of the early differentiation blockage. Therefore, the LAT 4YF mice are unable to lead or suggest the phenotype observed for the LAT<sup>Y136F</sup>, and LAT<sup>Y175F+Y195F+Y235F</sup> mice. Moreover, none of the results of the previous studies  
15 on cell lines suggests such a phenotype. Furthermore, the phenotype obtained in mice with the mutation Y136F could not be extrapolated in order to deduce the expected phenotype of mice having a composite mutation Y175F+Y195F+Y235F because of the different effects of the mutation Y136F and the  
20 mutations Y175F, Y195F, and Y235F observed during the cell line studies.

The object of the present invention is to provide non-human animals having a mutated LAT gene of the invention leading  
25 to an exaggerated Th2 cell differentiation. By "gene" is intended cDNA or genomic sequence coding for a LAT protein. By "mutated LAT gene of the invention" is intended a LAT gene coding for a mutant LAT protein, the sequence of which corresponds to a wild type sequence and contains the  
30 mutation of a single tyrosine among the four distal ones corresponding to Y136 in the mouse LAT protein or the composite mutation of the three distal tyrosine residues (corresponding to Y175, Y195 and Y235 in the mouse LAT protein). For example, the tyrosine corresponding to Y136 in  
35 the mouse LAT protein is the residue Y132 in the human LAT protein. In a first preferred embodiment, said LAT gene coding for a mutant LAT protein contains a single mutation

of the tyrosine residue corresponding to Y136 in the mouse LAT protein. In a second preferred embodiment, said LAT gene coding for a mutant LAT protein contains the composite mutation of the three distal tyrosines, those corresponding 5 to Y175, Y195 and Y235 in the mouse LAT protein. Preferably, said non-human animals are mice and said non-human animals have the mutated gene coding for a mutant LAT protein, the sequence of which corresponds to a wild type sequence and contains the single mutation of the tyrosine residue at 10 position 136 or the composite mutation of the three distal tyrosine residues at positions 175, 195 and 235. Preferably, said mutation consists in the replacement by a residue preventing the association of the "tyrosine-based" sequences with the SH2 domain of proteins. More preferably, said 15 mutation consists in the replacement of the tyrosine by a phenylalanine (Y-F), an aspartic acid (Y-D) or a glutamic acid (Y-E). Still more preferably, said mutation consists in the replacement of the tyrosine by a phenylalanine (Y-F). Preferably, said non-human animals according to the 20 invention are mammals, and in particular, they are rodents. More preferably, said rodents are mice. Preferably, said animals are homozygous for the mutated LAT gene or are carrying a null allele of the LAT gene. Preferably, said mutated LAT gene is incorporated into the animal genome by 25 targeted insertion in order to keep said mutated LAT gene under the control of regulatory regions of the endogenous LAT gene.

By "distal" is intended the C-terminal end of the protein. 30 Therefore, the distal tyrosine residues are the tyrosines residues located at the C-terminal end of the protein.

In particular, the invention concerns any germ cell and somatic cell from said animals or any progeny thereof 35 containing the mutated LAT gene of the invention. More particularly, germ cells and somatic cells of said animals contain the mutated LAT gene of the invention as a result of

chromosomal incorporation into the animal genome, or into an ancestor of said animal. Preferably, said mutated LAT gene is incorporated into the animal genome by targeted insertion (homologous recombination) in order to keep said mutated LAT gene under the control of regulatory regions of the endogenous LAT gene.

Therefore, a further object of the invention is to provide a mutated mouse gene coding for a mutant LAT protein, the sequence of which corresponds to a wild type sequence and contains the single mutation of the tyrosine Y136, or a composite mutation of the tyrosine residues at positions 175, 195, and 235. Said mutation consists in the replacement by a residue preventing the association of the "tyrosine-based" sequences with the SH2 domain of proteins. Preferably, said mutation of the tyrosine leads to its replacement by a phenylalanine, an aspartic acid or a glutamic acid. More preferably, said mutation of the tyrosine leads to its replacement by a phenylalanine. In a preferred embodiment, the sequence of the gene encoding mutated mouse LAT<sup>Y136F</sup> protein corresponds to sequence ID N°1. The invention further includes chromosomal DNAs containing exon 7 of the mutated gene (SEQ ID N°2). The invention concerns a mouse containing said mutated mouse gene.

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The present invention also encompasses plasmids comprising a DNA or a part thereof, having a sequence corresponding to the mutated gene according to the invention. In a preferred embodiment, the plasmids of the invention contain a restriction enzyme cleavage site, which is introduced in the intron 3' of exon 7. Advantageously, the restriction enzyme cleavage site is a Bgl II restriction site.

Said plasmids are useful for the generation of non-human animals according to the present invention.

Consequently, the invention also encompasses non-human

embryos introduced with the plasmids of the invention, and non-human embryos obtained by homologous recombination using the plasmids of the invention. In a preferred embodiment, the non-human embryos are embryonic stem cells derived from 5 a mouse. Advantageously, the ES cells are CK35 129/SV ES cells.

The invention also concerns the LAT mutant murine protein sequence containing the single mutation of the tyrosine Y136 10 or a composite mutation of the tyrosine residues at positions 175, 195, and 235. Said mutation consists in the replacement by a residue preventing the association of the "tyrosine-based" sequences with the SH2 domain of proteins. Preferably, said mutation of the tyrosine leads to its 15 replacement by a phenylalanine, an aspartic acid or a glutamic acid. More preferably, said mutation of the tyrosine leads to its replacement by a phenylalanine. In one embodiment, the invention concerns a mutated LAT protein containing the mutated amino acid sequence of exon 7 (SEQ ID 20 N°3).

The magnified and accelerated sequence of pathological events observed in the LAT <sup>Y136F</sup>, and LAT <sup>Y175F+Y195F+Y235F</sup> mice permits to readily start tests and studies. For example, 25 mutant LAT <sup>Y136F</sup> mice phenotype is achieved when they are 4 weeks old.

The mutant non-human animal according to the invention are useful in various applications of interest, in particular: 30 - to analyze the impact of drugs on the molecular mechanisms that lead to exacerbated IgE production as well as tissue eosinophilia, and - as a bioreactor allowing the dedicated production of IgE antibody of known specificity prior to or following a step 35 of humanization of the mutated LAT mouse (preferably LAT<sup>Y136F</sup> or LAT <sup>Y175F+Y195F+Y235F</sup> mouse).

Consequently, the present invention provides models of allergy, and/or asthma disease comprising animals according to the invention. In particular, the animals of the invention can be used as models of eosinophilia and/or T<sub>H</sub>2 cells deregulation, more particularly T<sub>H</sub>2 cells accumulation.

Therefore, the invention concerns the use of a mutant non-human animal according to the present invention as a model of allergy and/or asthma disease. The invention also 10 concerns the use of a mutant non-human animal according to the present invention as a model of eosinophilia. More generally, the invention concerns the use of a mutant non-human animal according to the present invention as a model of T<sub>H</sub>2 cells deregulation, more particularly a model of T<sub>H</sub>2 15 cells accumulation.

Due to the increased sensitivity of population, health difficulties such as asthma or allergies are more frequent. The animals according to the invention are suitable models 20 to help the research in these domains.

Accordingly, the present invention provides a method of screening for a drug for treatment of allergy, asthma and/or disease associated with T<sub>H</sub>2 cell deregulation or activity 25 comprising the step of subjecting the animals according to the invention, which are administered with the drug to a comparison with said animals, not administered with the drug.

30 More particularly, the invention concerns a method of screening of drugs for treatment of allergy, asthma and/or disease associated with T<sub>H</sub>2 cell deregulation or activity comprising the step of:

- 1) administering a candidate drug to a non-human animal 35 having a LAT gene coding for a mutant LAT protein according to the present invention;
- 2) evaluating the effect of said drug on the symptom or sign

of allergy, asthma and/or disease associated with  $T_{H2}$  cell deregulation or activity; and

3) selecting the drug that reduces said symptom or sign.

In a preferred embodiment, said screening method uses non-human animals not administered with drugs as control experiments. In an other preferred embodiment, said effect of said drug can be evaluated by measuring at least one parameter selected from the group : IgE level, IgG1 level, interleukin level (preferably IL-4, IL-10, IL-5 and/or IL-13), and eosinophilia. More preferably, said effect of said drug is evaluated by measuring the serum level of IgE and/or IgG1.

The invention also contemplates a method of screening drugs for treatment of allergy, asthma and/or disease associated with  $T_{H2}$  cell deregulation or activity comprising the step of:

1) subjecting cells having a LAT gene coding for a mutant LAT protein according to the present invention to a candidate drug;

2) evaluating the effect of said drug on said cells;

3) selecting the drug having the desired effect.

In a preferred embodiment, said effect of said drug can be evaluated by measuring the interleukin production, more particularly the IL-4 production.

An other object of this invention resides in a method of screening drugs that regulate the activity of  $T_{H2}$  cells, comprising the step of:

1) administering a candidate drug to a non-human animal having a LAT gene coding for a mutant LAT protein according to the present invention; and

2) selecting a drug that modulates the activity of  $T_{H2}$  cells in said non-human animal.

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The screening methods can be used to select, identify, characterize and/or optimize candidate drugs. The candidate

drugs may be of any origin, nature and structure. Their concentration may be adjusted by the skilled artisan. Furthermore, several drugs may be tested in parallel, or in combination.

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A further object of this invention is a method of producing a pharmaceutical composition for treating a disease associated with deregulated TH2 cells activity, particularly asthma or allergy, the method comprising (i) selecting, 10 identifying, optimizing or characterizing a compound using a screening assay as described above and (ii) conditioning said compound, or a derivative thereof, in a pharmaceutically acceptable carrier or vehicle.

15 In still another application, the present invention provides bioreactors for a large-scale production of human IgE antibodies comprising the animals according to the invention. LAT<sup>Y136F</sup> and LAT<sup>Y175F+Y195F+Y235F</sup> mice are indeed able to produce tremendous amount of IgE, as it is described in 20 example 2. IgE produced by mutant mice are useful for applications such as desensitization or for kit of clinical assay.

Therefore, the invention concerns a method of production of 25 human IgE antibodies comprising the steps of :

- 1) providing a non-human animal expressing humanized IgE;
- 2) breeding said animal expressing humanized IgE with a non-human animal having a LAT gene coding for a mutant LAT protein according to the present invention;
- 30 3) immunizing the animal of the progeny with an allergen;
- 4) recovering humanized IgE specific to said allergen.

The step 4 can comprise the step of producing B cell hybridomas producing said humanized IgE specific to said allergen. The invention relates to said B cell hybridoma 35 producing said humanized IgE specific to said allergen.

Said non-human animal expressing humanized IgE can be

obtained by conventional knock-in in which the genetic segment corresponding to the constant exons of the IgE gene is substituted by the corresponding human sequence.

5 The invention concerns the non-human animal resulting from the breeding of the animal expressing humanized IgE with the non-human animal having a LAT gene coding for a mutant LAT protein according to the present invention.

10 The produced humanized IgE specific to said allergen can be used for desensitization and in clinical assays aiming at characterizing allergens, preferably atopic allergens, present in patient.

15 The invention contemplates the oligonucleotide probes specific to a mutated LAT gene coding for a mutant LAT protein containing the single mutation of the tyrosine corresponding to Y136 in the mouse LAT protein or a composite mutation of the three distal tyrosines  
20 (corresponding to Y175, Y195 and Y235 in the mouse LAT protein). Such probes are useful to detect the presence of the mutation in a LAT gene. Hence, the invention provide oligonucleotides, the sequence of which corresponds to SEQ ID N°4 and/or SEQ ID N°5 as probes to screen the presence of  
25 the mutation Y136 in the mouse LAT gene. More particularly, the invention concerns oligonucleotide probes specific to a mutated human LAT gene coding for a mutant LAT protein containing a single mutation of the tyrosine Y132 or a composite mutation of the tyrosine residues Y171, Y191 and  
30 Y226. Such probes are useful for the detection of mutant LAT gene involved in asthma, allergy, eosinophilia and/or any disease associated with a Th2 cells deregulation or activity. Said probes can be part of a diagnostic kit.

35 Therefore, the invention relates to a diagnostic method for asthma, allergy, eosinophilia and/or Th2 cells deregulation, more particularly Th2 cells accumulation, comprising the

detection of a mutated LAT gene coding for a mutant LAT protein containing a single mutation of Y132 or a composite mutation Y171+Y191+Y226. Additionally, the invention concerns a diagnostic kit for asthma, allergy, eosinophilia and/or Th2 cells deregulation, more particularly Th2 cells accumulation, comprising oligonucleotide probes for the detection of a mutated LAT gene coding for a mutant LAT protein containing a single mutation of Y132 or a composite mutation Y171+Y191+Y226.

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Other characteristics and advantages of the invention are given in the following examples with reference to figures 1 to 11.

15 **EXAMPLES****Mutation LAT<sup>Y136</sup>****Example 1: Production of mutant mice**

To test *in vivo* the importance of LAT<sup>Y136</sup>, the inventors 20 generated knock-in mice with a mutation replacing Y136 with phenylalanine (Y136F).

**1. Materials and methods**25 **Mice**

Mice were maintained in a specific pathogen-free animal facility.

**LAT<sup>Y136F</sup> mutation.**

30 LAT genomic clones were isolated from a 129/Ola phage library. After establishing the nucleotide sequence and the exon-intron structure of the LAT gene, the tyrosine residue found at position 136 and encoded by exon 7 was mutated to phenylalanine. Mutagenesis was performed on a 1717-bp Eco 35 RI-Xba I fragment encompassing part of exon 5, exons 6, 7 and 8. In addition to the intended mutation, a new Bgl II

restriction enzyme cleavage site was introduced in the intron 3' of exon 7 to accommodate the LoxP-flanked *neo*<sup>r</sup> gene and facilitate subsequent identification of LAT<sup>Y136F</sup> mutant mice. Finally, the targeting construct was extended to give 5 1.7 kb and 4.8 kb of homologous sequences 5' and 3' of the EcoRI-XbaI fragment, respectively (see Fig. 2). After electroporation of CK35 129/SV ES cells (C. Kress et al., 1998), and selection in G418, colonies were screened for homologous recombination by Southern blot analysis. The 5' 10 single-copy probe is a 0.9-kb Bgl II-Xba I fragment isolated from a LAT genomic clone. When tested on Bgl II-digested DNA, the 5' probe hybridizes either to a 8.5 kb wild-type fragment or to a 4.5 kb recombinant fragment. Homologous recombination events at the 3' side were screened by long 15 range PCR. Homologous recombinant ES clones were further checked for the presence of the intended mutation by sequencing the genomic segment corresponding to exon 7. Finally, a *neo* probe was used to ensure that adventitious non homologous recombination events had not occurred in the 20 selected clones.

#### Production of mutant mice.

Mutant ES cells were injected into Balb/c blastocysts. Two LAT<sup>Y136F</sup> recombinant ES cell clones were found capable of germ 25 line transmission. The two mutant mouse lines were first bred to Deleter mice (Schwenk. F et al., 1995) to eliminate the Lox P-flanked neomycin cassette, and intercrossed to produce homozygous mutant mice. The two independently-derived mutant lines showed indistinguishable phenotype. To 30 confirm that the LAT Y136F mutation had been genuinely introduced, LAT transcripts were cloned by reverse transcription and PCR amplification from the thymus of the mutated mice, and the presence of the intended mutation confirmed by DNA sequence analysis. Screening of mice for 35 the presence of the LAT Y136F mutation was performed by PCR using the following pairs of oligonucleotides:  
c : 5'-GTGGCAAGCTACGAGAACCGAGGT-3' (SEQ ID N°4);

d : 5'-GACGAAGGAGCAAAGGTGGAAGGA-3' (SEQ ID N°5).

The single Lox P site remaining in the LAT Y136F allele after deletion of the  $neo^r$  resulted in an amplified PCR product 140 bp-longer than the 510 bp-long fragment 5 amplified from the wild-type LAT allele.

## 2) Mutant mice development

Mice homozygous for the LAT<sup>Y136F</sup> mutation, hereafter denoted LAT<sup>Y136F</sup>, were born at expected Mendelian frequencies and 10 their T cells contained levels of LAT proteins similar to wild-type T cells. At birth LAT<sup>Y136F</sup> mice displayed peripheral lymphoid organs of normal size. Beginning at about 3 weeks, however, the spleen and lymph nodes of the mutant mice started to enlarge relative to wild-type littermates, such 15 that by 15 weeks of age, spleen cellularity was approximately 10 times that of wild-type mice (Fig.3 A-C). Despite marked lymphocytic infiltrations in the lung, liver and kidney, homozygotes lived to at least 17 weeks of age, and no chronic intestinal inflammation or tumor formation 20 was observed. The effects of the LATY136F mutation were only detectable after breeding mice to homozygosity or to mice carrying a null allele of the LAT gene.

## 25 Example 2: Effect of the mutation: spontaneous exaggerated T helper type 2 immunity in mice

### 1. Materials and methods

#### 30 Purification of CD4+ T cells and eosinophils.

Lymph node and spleen cells from several mice were pooled and the CD4+ cells purified using a high gradient magnetic cell separation system (S. Miltenyi et al., 1990). Eosinophils were sorted on a FACSvantage™ on the basis of 35 their FSChigh, HSA+, and CD11b+ phenotype.

**Antibodies and flow cytometric analysis.**

Before staining, cells were preincubated on ice for at least 10 min with polyclonal mouse and rat Ig to block Fc receptors. Flow cytometric analysis was performed as 5 described previously (M. Malissen et al., 1995). All the antibodies were from BD PharMingen except the anti-CCR3 antibody that was purchased from R&D.

**Staining for intracellular cytokines.**

10 Before intracellular cytokine staining, cells ( $1.5 \times 10^6$ ) were cultured for 4 h in the presence of monensin (GolgiStop; BD PharMingen) at a final concentration of 2  $\mu$ M. Cells were then immediately placed on ice, washed, resuspended in PBS 1X, 1% FCS, 0.20% sodium azide, and 15 stained with an APC-conjugated anti-CD4 antibody. For intracellular cytokine staining, cells were first fixed using the cytofix/cytoperm kit (BD PharMingen). Each cell sample was subsequently split into aliquots that were separately stained with (1) a combination of FITC-conjugated 20 anti-IFN- $\alpha$  and PE-conjugated anti-IL-2 antibodies, (2) a combination of FITC-conjugated anti-IL-5 and PE-conjugated anti-IL-4 antibodies, and (3) a combination of fluorochrome-conjugated and isotype-matched negative control Ig (BD PharMingen). After a final wash, CD4+ cells ( $10^4$ ) were 25 analyzed on a FACSCalibur™ flow cytometer after gating out dead cells using forward and side scatters.

**RNase protection assay.**

For multiplex cytokine transcript analysis, total cellular 30 RNA was isolated from the specified cells using TRIzol (GIBCO-BRL Life Technologies) and analyzed by ribonuclease protection assay using an MCK-1 RiboQuant™ custom mouse template set (BD Pharmingen). Briefly,  $^{32}$ P-labeled riboprobes were mixed with 10  $\mu$ g of RNA, incubated at 56°C for 12 to 16 35 hours, and then treated with a mixture of RNases A and T1 and proteinase K. RNase-protected  $^{32}$ P-labeled RNA fragments

were separated on denaturing polyacrylamide gels and the intensity of the bands evaluated with a Fuji imaging plate system.

5 **Determination of serum isotype-specific immunoglobulin levels.**

The titres of polyclonal IgM, IgG1, IgG2a, IgG2b, IgG3 and IgA antibodies and  $\kappa$  and  $\lambda$  light chains were determined using isotype-specific ELISA (Southern Biotechnology). The 10 concentrations of IgG1 and IgE were determined by comparing test sample dilution series values with isotype control standards.

**2. Results**

15 A prominent phenotype of the CD4 T cells found in LAT<sup>Y136F</sup> mice was revealed when the inventors measured their ability to make cytokines. Due to the short half-lives of cytokines and of their transcripts, their analysis generally requires restimulation of T cells *in vitro* with PMA and ionomycin. A 20 multiprobe RNase protection assay detecting levels of transcripts of 9 cytokines showed that CD4 T cells freshly isolated from LAT<sup>Y136F</sup> mice contained sufficient IL-4 and IL-10 transcripts to allow their detection even without *ex vivo* restimulation (Fig. 4A). Upon activation by PMA/ionomycin 25 the levels of IL-4 and IL-10 transcripts they contained were further increased, and IL-5, IL-13, and IFN- $\alpha$  transcripts became readily detectable (Fig. 4B). In marked contrast, wild-type CD4 T cells yielded only the IL-2 and IFN- $\alpha$  transcripts expected for primary T cells. Analysis of IL-4 30 production at the single cell level, showed that following a 4 hr activation with PMA/ionomycin, close to 80% of the CD4 T cells isolated from LAT<sup>Y136F</sup> mice expressed very high levels of IL-4 (Fig. 4C). Consistent with the notion that these CD4 T cells were refractory to TCR stimuli, none of them scored 35 as IL-4+ in response to TCR cross-linking (Fig. 4C). Thus, LAT Y136F spontaneously developed a high frequency of Th2

cells. In the case of wild-type CD4 T cells, Th2 polarization of such magnitude is only achieved following prolonged antigenic stimulation in the presence of IL-4.

5 Light scatter analysis of thymic and lymph node cells from LAT<sup>y136F</sup> mice older than 4 weeks revealed a unique cell population that was almost absent from age-matched wild-type mice, and showed both an intermediate forward scatter and a high side scatter (Fig. 5A, 5B, 6A). Based on several of 10 criteria, these cells were identified as eosinophils (Fig. 6). Minute numbers of eosinophils normally reside in wild-type thymi, and their augmentation in LAT<sup>y136F</sup> thymi may primarily result from an intrinsic expression of LAT<sup>y136F</sup> molecules. However, LAT transcripts were undetectable in 15 eosinophils purified from LAT<sup>y136F</sup> mice, meaning that the thymic and lymph node eosinophilia they manifest result from the production of IL-5 by the abnormal CD4 cells present in these mutant mice.

20 Secondary lymphoid organs of 6-week old LAT<sup>y136F</sup> mice contained 7 to 10 times more B cells than their wild-type counterparts. Thus, the splenomegaly and generalized lymphadenopathy that developed in young LAT<sup>y136F</sup> mice can be mostly accounted for by cells belonging to the T and B cell 25 lineages. Over 90% of the mature B cells found in the spleen and lymph nodes of 6-week old wild type littermates had a resting phenotype (Fig. 7A). In marked contrast, only 25% of the B cells found in the enlarged secondary lymphoid organs of age-matched LAT<sup>y136F</sup> littermates showed a resting 30 phenotype. Among the remaining B cells, 25% showed an hyperactivated phenotype, and 50% expressed a phenotype typical of antibody producing cells. Coincident with the presence of these latter cells, serum IgG1 concentrations were elevated approximately 200 times compared to wild-type 35 mice, whereas those of IgE were elevated 2500 to 10000 times (Fig. 7C). In contrast, the levels of the other Ig isotypes did not differ significantly from those of wild-type serum

(Fig. 7B). In support of a polyclonal hypergammaglobulinemia G1 and E, the concentrations of kappa and lambda light chains were both markedly augmented in the serum of LAT<sup>Y136F</sup> mice (Fig. 7B). Notably, IgE and IgG1 antibody concentrations reached a plateau as early as 5 weeks of age (Fig. 7C), the values of which exceeded the extraordinarily large amounts of IgE and IgG1 previously reported for mice deprived of NFATc2 and NFATc3 transcription factors. Given that B cells do not express LAT proteins, and considering that isotype switching to IgE and IgG1 is highly dependent on the presence of IL-4 and IL-13, the overproduction of IgE and IgG1 noted in LAT<sup>Y136F</sup> mice is secondary to the presence of an abnormally high frequency of T<sub>H</sub>2 effectors.

15 **Example 3: Production of IgE**

Mice expressing humanized IgE are developed by conventional knock-in strategy in which the genetic segment corresponding to the constant exons of the IgE gene is substituted by the corresponding human sequence. Mice with a humanized IgE locus are bred into LAT Y136F mice. Following immunization, B cell hybridomas producing specific human IgE are produced, and the resulting specific human IgE are used as "standard" in clinical assays aiming at characterizing atopic allergens present in patients."

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**Example 4: Screening for a drug**

Mutant mice and control ones will be treated with a variety of drugs or original compounds. Their effects will be analyzed *in vivo* by measuring various parameters such as:

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- T<sub>H</sub>2 cells differentiation.
- Production of T<sub>H</sub>2 types cytokines
- Eosinophilia
- Hypergammaglobulinemia G1 and E .

**Example 5: Production of mutant mice**

To test *in vivo* the importance of the three carboxy-terminal 5 tyrosines (LAT Y175, LAT Y195 and LAT Y235), the inventors generated knock-in mice with a mutation replacing these three tyrosines with phenylalanine (LAT Y175F + Y195F + Y235F).

10 **1. Materials and methods****Mice**

Mice were maintained in a specific pathogen-free animal facility.

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**LAT Y175F + Y195F + Y235F mutation.**

LAT genomic clones were isolated from a 129/Ola phage library. After establishing the nucleotide sequence and the exon-intron structure of the LAT gene (EMBL Nucleotide 20 Sequence Datatase; accession number: AJ438435), the tyrosine residues found at positions 175, 195 and 235 and encoded by exons 9, 10, 11 were mutated to phenylalanine. Mutagenesis was performed on a 815 bp Ncol-BamHI fragment encompassing exons 9, 10, 11 (coding for tyrosines 175 (exon 9), 195 25 (exon 10) and 235 (exon 11) and part of exon 12 (corresponding to the 3' untranslated region of LAT). Each exon was mutated independently and new restriction sites were introduced for facilitating subsequent cloning steps. A new Eco RI site was introduced on the 5' side of the Ncol site, BamHI and ClaI sites were introduced between exons 9 and 10, a HindIII site was introduced between exons 10 and 30 11, and BgIII, XhoI, and NotI sites were introduced on the 3' side of exon 11 in lieu of the original BamHI site. PCR reactions were performed with Pwo DNA polymerase (Boehringer 35 Mannheim), and PCR products were purified and cut with EcoRI and BamHI for exon 9, BamHI and HindIII for exon 10,

and HindIII and NotI for exon 11. These three fragments were assembled in a pBS-KS plasmid (Stratagene). The resulting plasmid was used to clone a 3.5 kb Eco RI-Nco I genomic fragment providing a 5' homology arm and a 4.3 kb-Sal I genomic fragment providing a 3' homology arm. Finally a loxP flanked neo<sup>r</sup> gene was introduced using the BamHI and ClaI sites that were engineered between exons 9 and 10. After electroporation of CK35 129/SV embryonic stem (ES) cells and selection in G418, colonies were screened for homologous recombination by southern blot analysis using a 3' single-copy probe that consisted of a 1.1 kb EcoRI-HindIII fragment isolated from a LAT genomic clone. When tested on BamHI digested genomic DNA, the 3' probe hybridizes either to a 7.0 kb wild-type fragment or to a 9.1 kb recombinant fragment. The presence of a genuine recombination event was checked by PCR using the following pair of primers (depicted in Figure 8):

**f:** 5'-CCCAGAGGCAAACCCCTCTGAAG-3' (SEQ ID N°6) and

**g:** 5'-TCGAATTCGCCAATGACAAGACGC-3' (SEQ ID N°7). This PCR gives a band of 8.6 kb in the recombinant ES clones only. Homologous recombinant ES clones were further checked for the presence of the intended mutations by sequencing the genomic segment corresponding to exons 9, 10 and 11. Finally, a neo probe was used to ensure that adventitious non-homologous recombination events had not occurred in the selected clones.

#### **Production of mutant mice.**

Mutant ES cells were injected into Balb/c blastocysts. Two LAT Y175F + Y195F + Y235F recombinant ES cell clones were found capable of germ line transmission. The two mutant mouse lines were first bred to Deleter mice (Schwenk. F et al., 1995) to eliminate the Lox P-flanked neomycin cassette, and intercrossed to produce homozygous mutant mice. The two independently-derived mutant lines showed indistinguishable phenotype. To confirm that the LAT Y175F + Y195F + Y235F mutation had been genuinely introduced, LAT transcripts were

cloned by reverse transcription and PCR amplification from the thymus of the mutated mice, and the presence of the intended mutation confirmed by DNA sequence analysis. Screening of mice for the presence of the LAT Y175F + Y195F + Y235F mutation was performed by PCR using the following pairs of oligonucleotides:

d: 5'-GGAGACTTAGATGTCTGAGCCG-3' (SEQ ID N°8) and  
e: 5'-GACAGACCAGCAGGGACAGTG-3' (SEQ ID N°9) (Wt 238bp, mutant 435bp).

10 The single Lox P site remaining in the LAT Y175F + Y195F + Y235F allele after deletion of the neo<sup>r</sup> resulted in an amplified PCR product 140 bp-longer than the 510 bp-long fragment amplified from the wild-type LAT allele.

15 **2) Mutant mice development**

Mice homozygous for the LAT Y175F + Y195F + Y235F mutation, hereafter denoted LAT Y175F + Y195F + Y235F were born at expected Mendelian frequencies and their T cells contained levels of LAT proteins similar to wild-type T cells. At 20 birth LAT Y175F + Y195F + Y235F mice displayed peripheral lymphoid organs of normal size. Beginning at about 3 months, however, the spleen and lymph nodes of the mutant mice started to enlarge relative to wild-type littermates, such that by 3 months of age, spleen cellularity was 25 approximately 5 times that of wild-type mice. Homozygotes lived to at least 5 months of age, and no chronic intestinal inflammation or tumor formation was observed. The effects of the LAT Y175F + Y195F + Y235F mutation were only detectable after breeding mice to homozygosity or to mice carrying a 30 null allele of the LAT gene.

**Example 6: Effect of the mutation: a subset of gamma/delta T cells expands and acquire a spontaneous exaggerated T helper type 2 immunity in mice.**

**1. Materials and methods**

**Purification of gamma/delta T cells and eosinophils.**

Spleen cells from several mice were pooled and the gamma/delta T cells purified using a high gradient magnetic cell separation system (S. Miltenyi et al., 1990).

**Antibodies and flow cytometric analysis.**

Before staining, cells were preincubated on ice for at least 10 min with polyclonal mouse and rat Ig to block Fc receptors. Flow cytometric analysis was performed as described previously (M. Malissen et al., 1995). All the antibodies were from BD PharMingen.

**Staining for intracellular cytokines.**

Before intracellular cytokine staining, cells ( $1.5 \times 10^6$ ) were cultured for 4 h in the presence of monensin (GolgiStop; BD PharMingen) at a final concentration of 2  $\mu$ M. Cells were then immediately placed on ice, washed, resuspended in PBS 1X, 1% FCS, 0.20% sodium azide, and stained with an APC-conjugated anti-CD5 antibody. For intracellular cytokine staining, cells were first fixed using the cytofix/cytoperm kit (BD PharMingen). Each cell sample was subsequently split into aliquots that were separately stained with (1) a combination of FITC-conjugated anti-IFN- $\alpha$  and PE-conjugated anti-IL-2 antibodies, (2) a combination of FITC-conjugated anti-IL-5 and PE-conjugated anti-IL-4 antibodies, and (3) a combination of fluorochrome-conjugated and isotype-matched negative control Ig (BD PharMingen). After a final wash, CD5+ cells ( $10^4$ ) were analyzed on a FACSCalibur™ flow cytometer after gating out dead cells using forward and side scatters.

**RNase protection assay.**

For multiplex cytokine transcript analysis, total cellular RNA was isolated from the specified cells using TRIzol 35 (GIBCO-BRL Life Technologies) and analyzed by ribonuclease protection assay using an MCK-1 RiboQuant™ custom mouse

template set (BD Pharmingen). Briefly,  $^{32}\text{P}$ -labeled riboprobes were mixed with 10  $\mu\text{g}$  of RNA, incubated at 56°C for 12 to 16 hours, and then treated with a mixture of RNases A and T1 and proteinase K. RNase-protected  $^{32}\text{P}$ -labeled RNA fragments were separated on denaturing polyacrylamide gels and the intensity of the bands evaluated with a Fuji imaging plate system.

**Determination of serum isotype-specific immunoglobulin levels.**

The titres of polyclonal IgM, IgG1, IgG2a, IgG2b, IgG3 and IgA antibodies and  $\square$  and  $\square$  light chains were determined using isotype-specific ELISA (Southern Biotechnology). The concentrations of IgG1 and IgE were determined by comparing test sample dilution series values with isotype control standards.

**2. Results**

A prominent phenotype of the CD90.2 $^+$ , CD5 $^+$  gamma/delta T cells found in LAT Y175F + Y195F + Y235F mice (see Figure 9) was revealed when the inventors measured their ability to make cytokines. Due to the short half-lives of cytokines and of their transcripts, their analysis generally requires restimulation of T cells *in vitro* with PMA and ionomycin. A multiprobe RNase protection assay detecting levels of transcripts of 9 cytokines showed that gamma/delta T cells freshly isolated from LAT Y175F + Y195F + Y235F mice contained large amounts of IL-4, IL-5, IL-10 and IL-13 transcripts to (Figure 10). This attribute is reminiscent of the observation made with the alpha/beta T cells present in the periphery of the LAT Y136F mice. In marked contrast, wild-type CD4 T cells yielded only the IL-2 and IFN- $\gamma$  transcripts expected for primary T cells. Analysis of IL-4 production at the single cell level, showed that following a 4 hr activation with PMA/ionomycin, close to 80% of the CD4 T cells isolated from LAT Y175F + Y195F + Y235F mice expressed very high levels of IL-4. Thus, LAT Y175F + Y195F

+ Y235F mice spontaneously developed a high frequency of gamma/delta T cells with a Th2 phenotype. In the case of wild-type CD4 T cells, Th2 polarization of such magnitude is only achieved following prolonged antigenic stimulation in 5 the presence of IL-4.

The spleen of 3-month old LAT Y175F + Y195F + Y235F mice contained 5 to 10 times more B cells than their wild-type counterparts. Thus, the splenomegaly that developed in LAT 10 Y175F + Y195F + Y235F mice can be mostly accounted for by cells belonging to the T and B cell lineages. Over 90% of the mature B cells found in the spleen and lymph nodes of 3-month old wild type littermates had a resting phenotype (Fig. 11A). In marked contrast, only 16% of the B cells 15 found in the enlarged secondary lymphoid organs of age-matched LAT Y175F + Y195F + Y235F littermates showed a resting phenotype. Among the remaining B cells, 21% showed an hyperactivated phenotype, and 63% expressed a phenotype typical of antibody producing cells. Coincident with the 20 presence of these latter cells, serum IgG1 concentrations were elevated approximately 100 times compared to wild-type mice, whereas those of IgE were elevated 500 to 5000 times (Fig. 11). In contrast, the levels of the other Ig isotypes did not differ significantly from those of wild-type serum. 25 Given that mature B cells do not express LAT proteins, and considering that isotype switching to IgE and IgG1 is highly dependent on the presence of IL-4 and IL-13, the overproduction of IgE and IgG1 noted in LAT Y175F + Y195F + Y235F mice is secondary to the presence of an abnormally 30 high frequency of gamma/delta T cells producing Th2 cytokines.

#### Example 7: Production of IgE

Mice expressing humanized IgE are developed by conventional 35 knock-in strategy in which the genetic segment corresponding to the constant exons of the IgE gene is substituted by the corresponding human sequence. Mice with a humanized IgE

locus are bred into LAT Y175F + Y195F + Y235F mice. Following immunization, B cell hybridomas producing specific human IgE are produced, and the resulting specific human IgE are used as "standard" in clinical assays aiming at 5 characterizing atopic allergens present in patients."

**Example 8: Screening for a drug**

Mutant mice and control ones will be treated with a variety of drugs or original compounds. Their effects will be 10 analyzed in vivo by measuring various parameters such as:

- Th2 cells differentiation.
- Production of Th2 types cytokines
- Hypergammaglobulinemia G1 and E .

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